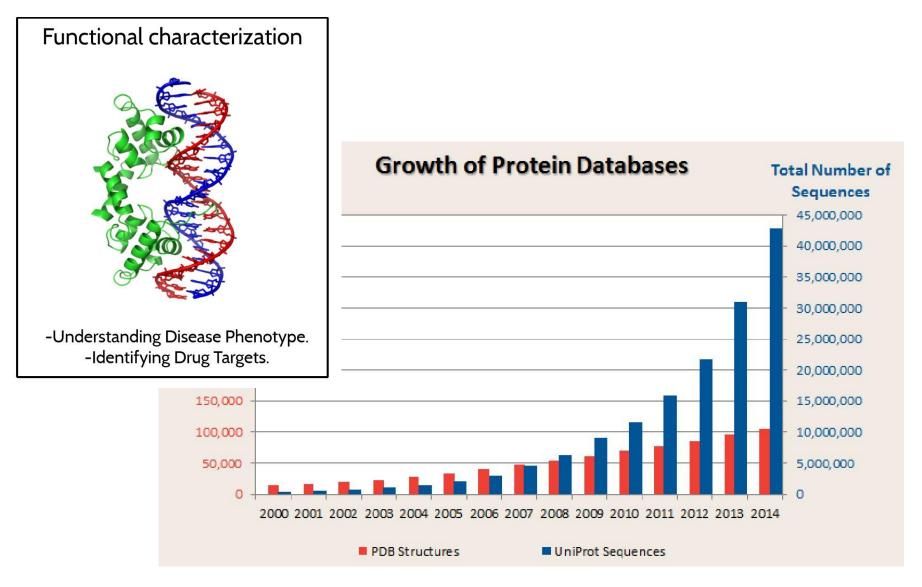
# Biologically inspired de novo protein structure prediction

Charlotte Deane
Department of Statistics
Oxford University

# Why predict protein structures?



## Structure prediction methods

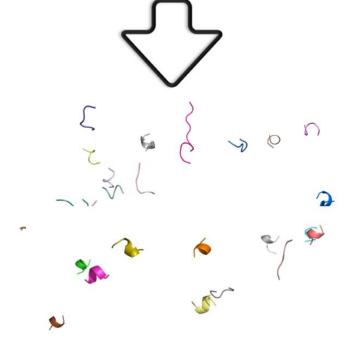
- Template-based methods:
  - Comparative modelling (or Homology modelling):
    - There exists a protein with clear homology.
    - Uses sequence-based techniques to identify a template. – Protein Threading/Fold recognition:
    - There exists a protein of similar fold (analogy).
- Template-free methods:
  - Novel fold prediction

# Fragment assembly – Protein structure prediction

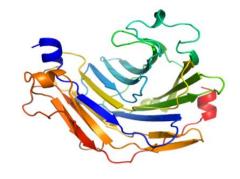
RPRTAFSSEQLARLKREFNENR YLTERRRQQLSSELGLNEAQIKI WFQNKRAKI











# Fragment assembly – Protein structure prediction



Where for any given position, there are multiple pieces that can fit in it...



Where the pieces got mixed up with pieces from another puzzle...

Where some pieces are missing...

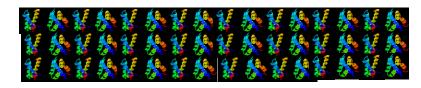


And where you cannot look at the box to check how it is supposed to look like...

#### How does it work?



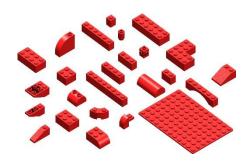
- Energy function
  - Usually from a Bayesian treatment of residue distributions in known protein structures sometimes combined with physics based energy terms
  - Pair potential terms, Solvation potentials terms, Steric terms, Longrange hydrogen bonding, compactness term
  - Predicted contacts from co-evolution methods
- Use a Monte Carlo search procedure
  - Move set based on fragments of protein structures
- Generate thousands of decoys



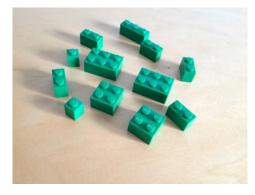
Select a final answer

Consider secondary structure when assessing your fragment library

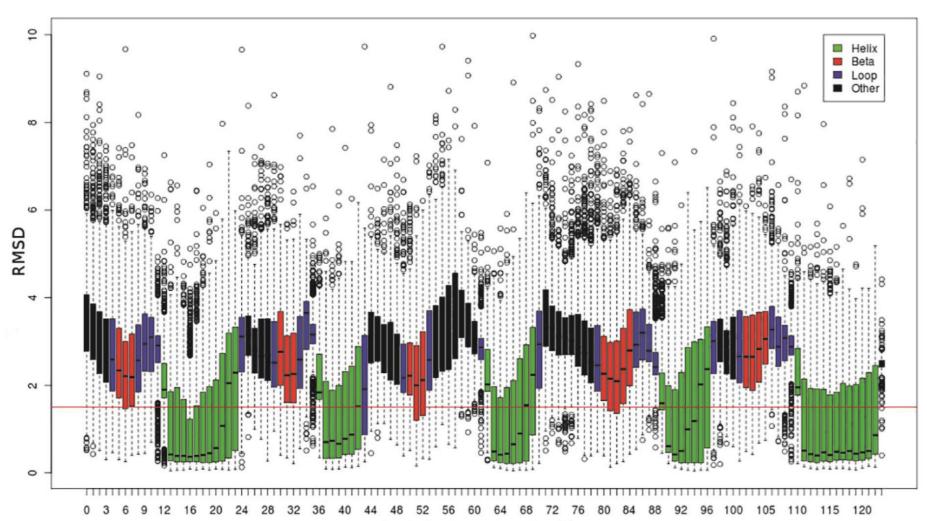
DREFGWTYPACDEFLMNGHIKLMNPQRSTVWY.....



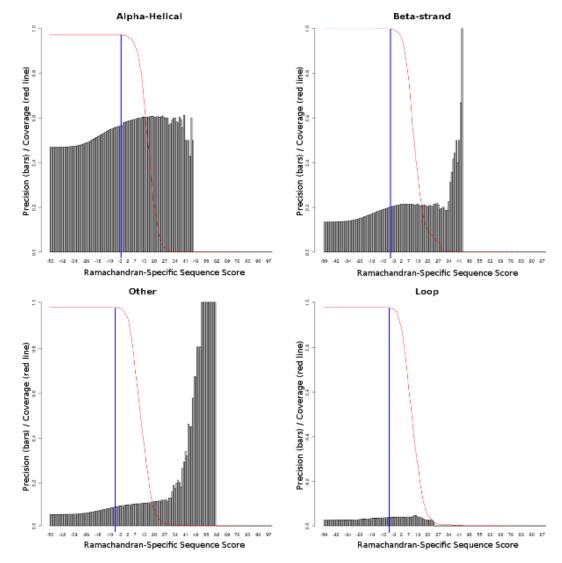




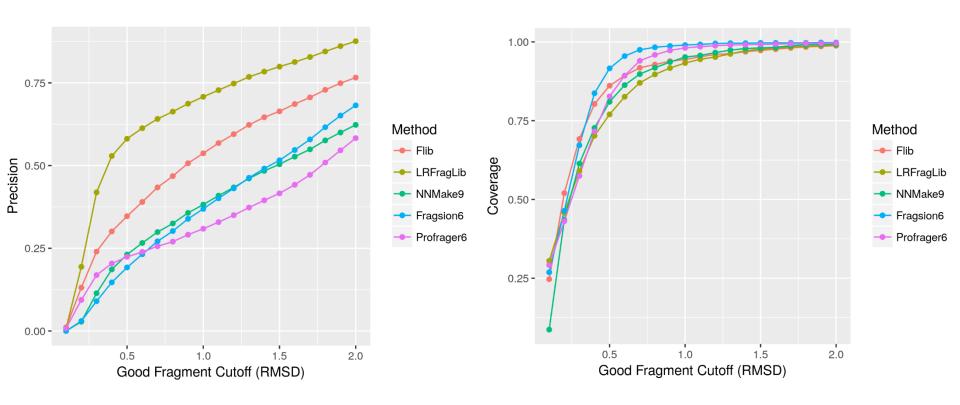
Consider secondary structure when assessing your fragment library



Consider secondary structure when assessing your fragment library



Consider secondary structure when assessing your fragment library



NNMAKE – Gront et al (2011)

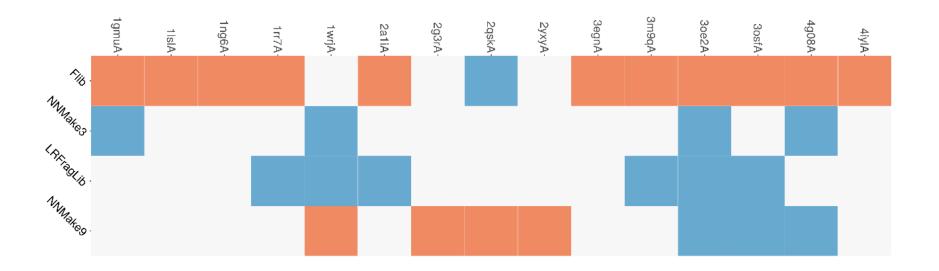
FLIB – Oliveira et al (2015)

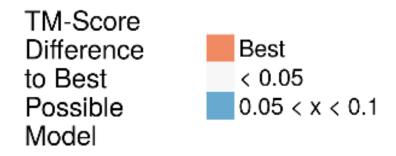
LRFragLib – Wang et al (2016)

Fragsion – Bhattacharya et al (2016)

Profrager – Santos et al (2015)

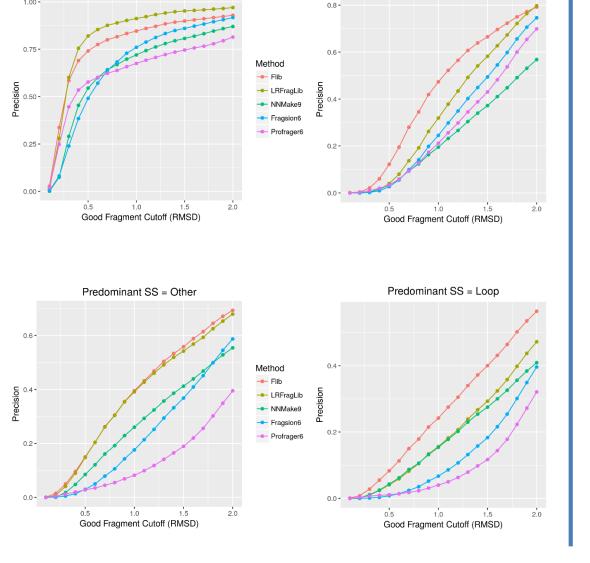
Consider secondary structure when assessing your fragment library





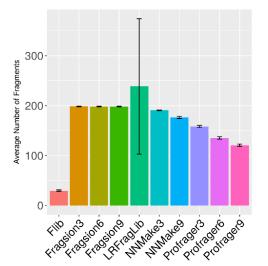
Consider secondary structure when assessing your fragment library

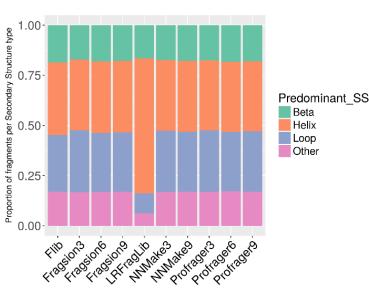
Predominant SS = Beta



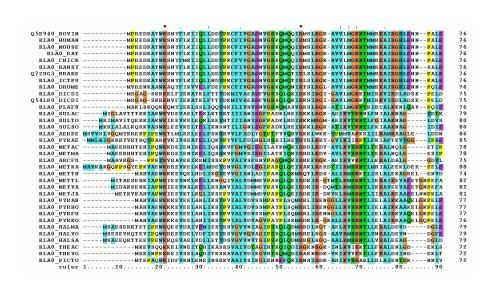
Predominant SS = Helix

1.00 -





### Looking for inspiration in biology: coevolution



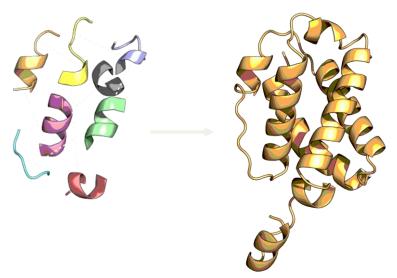
Multiple sequence alignments can be used to identify pairs of residues that coevolve.

Protein contacts can be predicted using this coevolutionary signal.

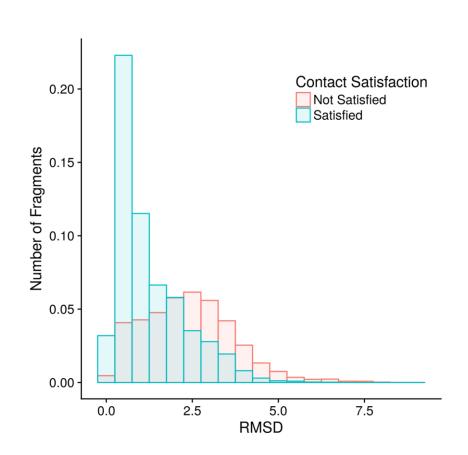


# Coevolution can be used to produce better fragment libraries

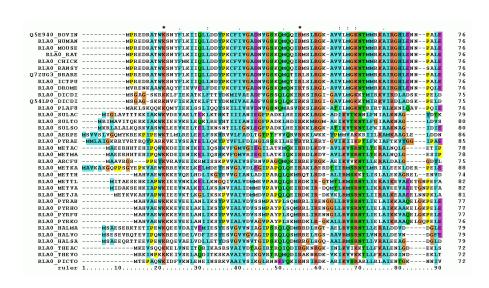
Fragments from known protein structures can be used to guide the conformational search.



Fragments that satisfy predicted contacts tend to be of better quality.

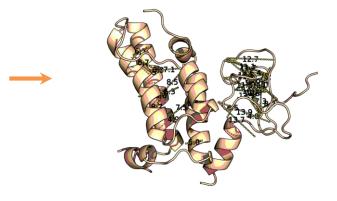


# Coevolution can be used in the energy function



Multiple sequence alignments can be used to identify pairs of residues that coevolve.

Protein contacts can be predicted using this coevolutionary signal.



Predicted contacts improve scoring and lead to accurate protein structure prediction.

#### Co-evolution methods

- Test set 3458 proteins
- FreeContact
- PSICOV
- CCMPred
- Bbcontacts
- metaPSICOV stage 1
- metaPSICOV stage2
- metaPSICOV HB
- GREMLIN

Kajan, L. et al. (2014)

Jones, D.T. et al. (2012)

Seemayer, S. et al. (2014)

Andreani and Soding (2015)

Jones, D.T. et al. (2014)

Jones, D.T. et al. (2014)

Jones, D.T. et al. (2014)

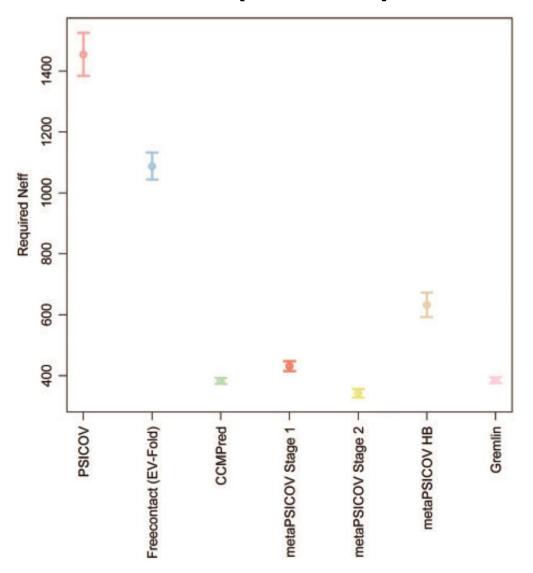
Kamisetty et al. (2013)

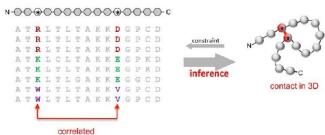
#### Contact definition

• Two protein residues are defined to be in contact if their C- $\beta$ s (C- $\alpha$ s for Glycine) are less than 8 A apart

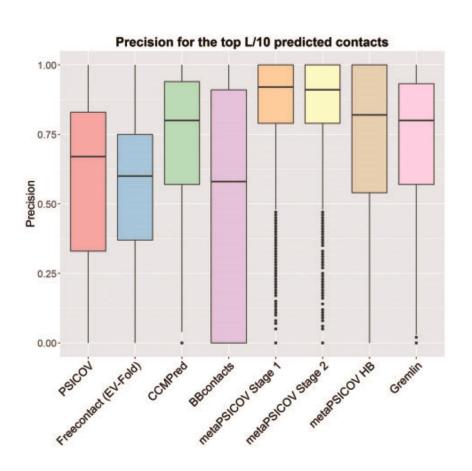
- Contacts between residues being less than five residues apart and are not considered
- A short-range contact between residues i and j is defined when 5 ≤ |i - j |≥23.
- A long range contact is defined when |i-j| > 23

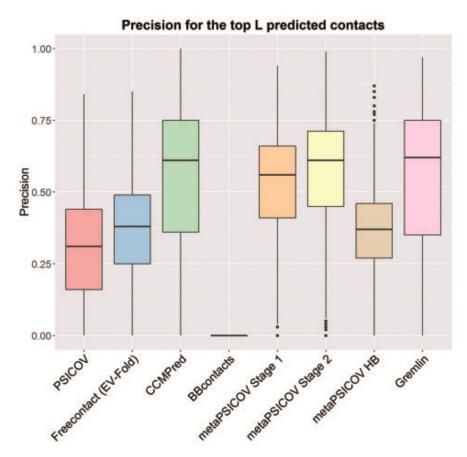
# How many sequences do you need in the multiple sequence alignment?





#### How accurate are the methods?





# Putting co-evolutionary contacts into protein structure prediction

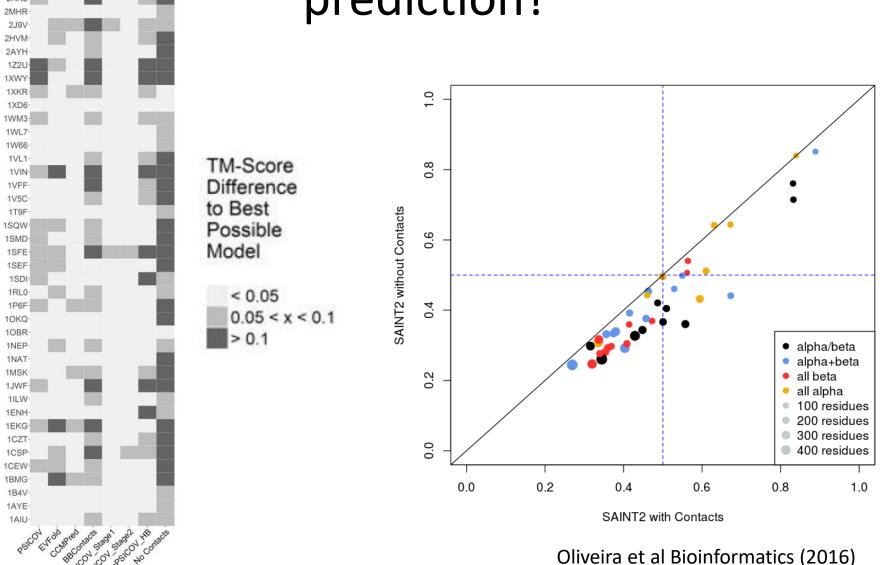
$$S_{ij}^{contact} = \begin{cases} 0, & \text{if } ||\mathbf{C}_{\beta}(i) - \mathbf{C}_{\beta}(j)|| < 8.0 \text{ Å} \\ ||\mathbf{C}_{\beta}(i) - \mathbf{C}_{\beta}(j)|| - 8.0 \text{ Å}, & \text{otherwise.} \end{cases}$$

Where  $C_{\beta}(i)$  and  $C_{\beta}(j)$  represent the coordinates of the C- $\beta$ s (C- $\alpha$ s in the case of glycine) of residues i and j and:

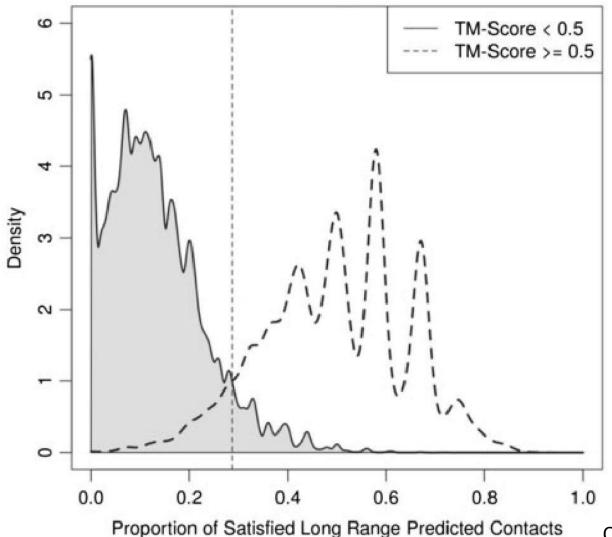
$$||\mathbf{C}_{\beta}(i) - \mathbf{C}_{\beta}(j)|| = \sqrt{\sum_{\kappa=x,y,z} (C_{\beta}^{\kappa}(i) - C_{\beta}^{\kappa}(j))^2}$$

# How do they influence structure prediction?

2YVT



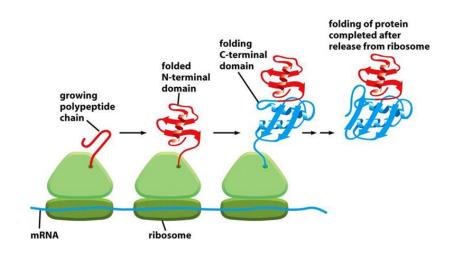
# Using co-evolution contacts to identify good models



# Inspiration from biology: cotranslational protein folding

 Folding is orders of magnitude faster than translation.

 Cotranslational protein folding not necessary for all proteins to reach their native state.



 Cotranslational protein folding is faster/more efficient than in vitro re-folding. Hypothesis: CT folding guides proteins towards their native state by restricting the conformational search space.

# Template-free search strategies with SAINT2

Co-translational, series of smaller optimisation problems
Therefore- faster



Traditional:
Non-sequential approach

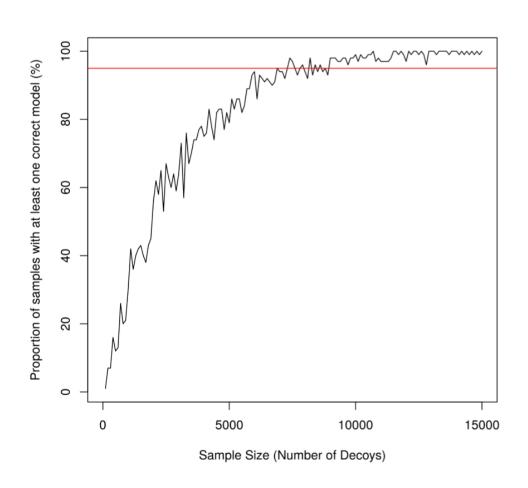
Biologically Inspired: Cotranslational approach

# Number of decoys required

Table 1. Number of decoys produced by different de novo structure predictors as described in recent works.

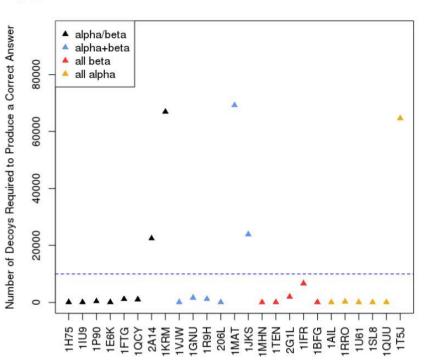
Method:	<b>Number of Decoys:</b>
FRAGFOLD (6)	200
CABS(7)	360
MBS (8)	3,000
RBOaleph (9)	1,000-5,000
<b>QUARK</b> (10)	5,000
Nefilim (11)	150,000
EDAfold (12)	200,000
Rosetta (13)	20,000-900,000

# Number of decoys required



# Number of decoys required

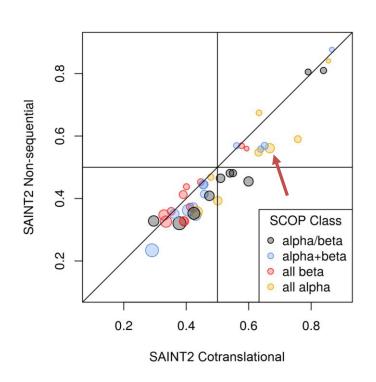


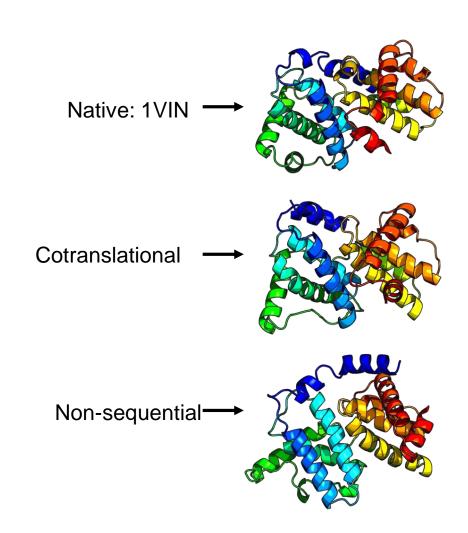


- Number decoys to get a correct answer ~10,000
- Number of decoys to get best answer ~20,000
- Not dependent on protein length (if length <250)</li>

# Cotranslational prediction produces better models

#### Comparing search strategies:

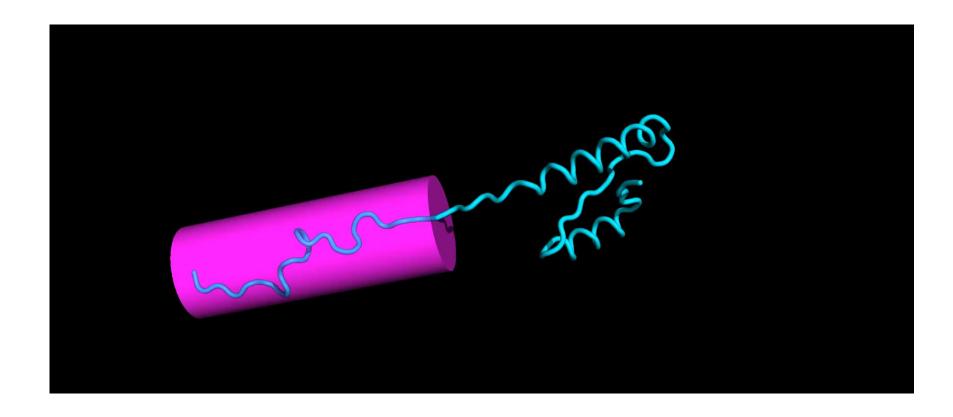




# Improving the search: Cotranslational protein structure prediction

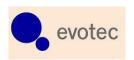
- Most current de novo structure prediction methods randomly sample protein conformations
  - Require large amounts of computational resource
- SAINT2 uses a sequential sampling strategy, suggested by biology
  - Requires fewer decoys to produce a good answer
- Sequential sampling improves speed
  - 1.5 to 2.5 times faster than non-sequential prediction.
- SAINT2 sequential produces better models
- SAINT2 sequential a pseudo-greedy search strategy that reduces computational time of de novo protein structure prediction and improves accuracy

## What next?



#### **ACKNOWLEDGEMENTS**

















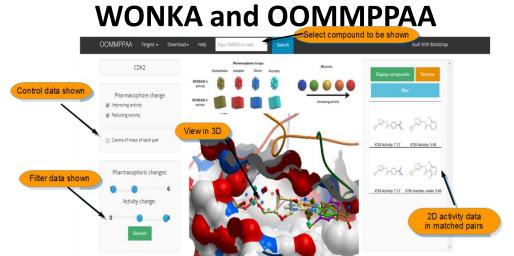














**Memoir** is a homology modelling algorithm designed for membrane proteins. The inputs are the sequence which is to be modelled, and the 3D structure of a template membrane protein. We have a short **video tutorial** on how to use Memoir and an **example results page**. We also have a tutorial on how to **model multiple chain transmembrane proteins**.

#### http://www.stats.ox.ac.uk/proteins/resources

